Benzo[b]cyclohept[e][1,4]oxazines§ IV.¹¹ Synthesis and Properties of All Possible Benzoxazinotropones²¹

Hidetsugu Wakabayashi,* Teruo Kurihara,* Sumio Ishikawa, Joji Okada,† and Tetsuo Nozoe††

Department of Chemistry, Faculty of Science, Josai University,

1-1 Keyakidai, Sakado-shi, Saitama 350-02

† Tochigi Research Laboratories, Kao Corporation,

2606, Akabane, Ichikaimachi, Tochigi 321-34

†† Tokyo Research Laboratories, Kao Corporation,

2-1-3 Bunka, Sumida-ku, Tokyo 131

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Five isomeric benzo[b]cyclohept[e][1,4]oxazin-6~10-ones (hereafter refered to as benzoxazinotropones) and bromobenzoxazinotropones were prepared almost quantitatively by the hydrolysis of the corresponding mono-, di-, and tribromobenzo[b]cyclohept[e][1,4]oxazines with refluxing acetic acid. On the basis of spectral data and theoretical calculations, benzo[b]cyclohept[e][1,4]oxazin-6~9(11H)-ones are believed to exist in the keto form. These compounds gave respective methoxy- and acetoxybenzo[b]cyclohept[e][1,4]oxazines when reacted with diazomethane and acetic anhydride. On the other hand, benzo[b]cyclohept[e][1,4]oxazin-10(11H)-one was found to exist in a stable, intermolecularly hydrogen-bonded form and did not form acetoxylated or methylated compounds. Possible reaction pathways for the formation of these products are discussed.

One of the authors (T.N.) and his co-worker reported³⁾ that a small amount (less than 1%) of isomeric isopropylbenzo[b]cyclohept[e][1,4]oxazin-10(11H)-ones (1a—c) were produced along with the main products 7-, 8-, and 9-isopropylbenzo[b]cyclohept[e][1,4]oxazines (2a—c), when isomeric 2-chloro-4-, 5-, and 6-isopropyltropones (3a—c) were refluxed in acetic acid with o-aminophenol (4). We later reported⁴⁾ that the con-

densation of 3-bromo-2-methoxytropone (5a) with an excess of 4 in refluxing acetic acid gave a small amount of

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benzo[b] cyclohept[e][1,4]oxazin-6(11H)-one (6), and its 10(11H)-one isomer (10) as 1:1-condensation products, along with various interesting 1:2-condensation products. Although some possible reaction pathways for the formation of 6 and 10 (and for 1) were presented, we did not have enough experimental evidence to distinguish between them. 1) In this paper we wish to

describe the details of the synthesis, structures, and keto-enol tautomerism of all possible isomers of the benzo-[b]cyclohept[e][1,4]oxazin-6~10(11H)-ones (hereafter referred to as benzoxazinotropones, 6—10), as well as a possible mechanism for the formation of these compounds.

Results and Discussion

Synthesis of Various Bromobenzo[b]cyclohept[e]-[1,4]oxazines (11a—e). 6-Bromo- $(11a)^{5}$) and 8-bromobenzo[b]cyclohept[e][1,4]oxazine (11c)⁶) were respectively prepared by the condensation of 7-bromo-(5d) and 5-bromo-2-methoxytropone (5c) with 4, via the respective bromo-2-(2-hydroxyanilino)tropones (12a, b). The 10-bromo compound (11e) was obtained from 11a by an intermolecular heterocycle exchange reaction⁷) with 4 in

[§] Benzo[b]cyclohept[e][1,4]oxazine was referred to as cyclohepta[b][1,4]benzoxazine in the preceding paper.

methanol (Scheme 1).

The reaction of 5-bromotropolone p-toluenesulfonate (14) with 4 in pyridine gave anilinotropone 12c (84% yield) through cine substitution.⁸⁾ Compound 12c underwent cyclization in hot acetic acid containing a trace amount of concd sulfuric acid to give 11d (82% yield, Scheme 2). The structures of 12c and 11d were determined to be 4-bromo-2-(2-hydroxyanilino)tropone and 9-bromobenzo[b]cyclohept[e][1,4]oxazine, respectively, on the basis of the spectral data (see the Experimental section).

The reaction of 11d with 2 equiv of 4 in MeOH at room temperature was studied by time-dependent HPLC. It

Scheme 1.

Scheme 2.

was determined that **11d** and **11b** attained an approximately 1:1 equilibrium within 1 h by an intermolecular heterocycle exchange reaction (Scheme 2).⁷⁾ The reaction mixture was separated by silica-gel chromatography and **11b** (45% yield) was identified as 7-bromobenzo[b]cyclohept[e][1,4]oxazine on the basis of NMR spectra and comparison of its UV spectra with **11d**.

Synthesis of Benzoxazinotropones. Heating of bromo compounds 11a-e in acetic acid at $110 \,^{\circ}$ C for 20 min to 2 h (depending on the position of the Br atom) gave the corresponding benzoxazinotropones (6—10) in nearly quantitative yields. Compounds 6 and 10 were identified as benzo[b]cyclohept[e][1,4]oxazin-6(11H)-one and -10(11H)-one, respectively, by direct comparison with the authentic samples.⁴⁾ The structure of 7 was shown spectroscopically to be benzo[b]cyclohept[e][1,4]oxazin-7(11H)-one. Compounds 8 and 9 were also shown to be benzo[b]cyclohept[e][1,4]oxazin-8(11H)-one and -9(11H)-one, respectively, on the basis of the spectral data (see the Experimental section).

Similarly, 6.8-dibromobenzo[b]cyclohept[e][1,4]oxazine (15)6) produced two bromobenzoxazinotropones (16 (5% yield) and 17 (80% yield)) as well as a small amount of 12d. The structures of 16 and 17 were determined to be 8-bromobenzo[b]cyclohept[e][1,4]oxazin-6-(11H)-one and 6-bromobenzo[b]cyclohept[e]-[1,4]oxazin-8(11H)-one, respectively, on the basis of high resolution mass spectral data and comparison of their UV spectra with those of 6 and 8, as well as the results of acylation (see below). On the other hand, 6,8,10-tribromobenzo [b] cyclohept [e] [1,4] oxazine $(18)^{9)}$ gave, under similar conditions, isomeric dibromobenzoxazinotropones 19 (60% yield) and 20 (20% yield). Zinc dust reduction of compounds 19 and 20 resulted in the same benzoxazinotropone 10. Compound 19 was also prepared by the reaction of 2,4,6-tribromo-7-methoxytropone (21) with 4. This evidence along with the spectral data mentioned above showed the structures of 19 and 20 to be 7,9-dibromo- and 6,8-dibromobenzo-[b]cyclohept[e][1,4]oxazin-10(11H)-one, respectively (Scheme 3).

Hydrolysis of 18 with AcOH gave only the dibromo derivatives of benzo[b]cyclohept[e][1,4]oxazin-10(11H)-one (10), but neither 6(11H)-one nor 8(11H)-one deriva-

Scheme 3.

tives were produced, whereas the same treatment of 15 gave a mixture of 6(11H)-one derivative 16 and 8(11H)-one derivative 17 in a 1:16 ratio. These results suggest that the facility of the hydrolytic cleavage is in order C-10>C-8>C-6.

The bonding energies between the Br-attached carbon and Br of 11a—e were calculated by the MINDO/3 method to be -11.38, -11.27, -11.32, -11.31, and -11.34 eV, respectively. In the case of 15, the binding energy between C-6 and Br was calculated to be -11.45 eV, while that between C-8 and Br was -11.35 eV. Therefore, the bond between C-8 and its Br atom is hydrolyzed more readily than the bond between C-6 and its Br atom. In the case of 18, the binding energies between a Br-attached carbon and its Br atom are in order C-6 (-11.47 eV) < C-8 (-11.42 eV) < C-10 (-11.40 eV). These results agree with the order of the facility of the hydrolytic cleavage.

Keto-Enol Tautomerism of Benzoxazinotropones. Keto-enol tautomerism is believed to exist in compounds 6—10. In order to clarify the tautomerism, the acetylation and methylation of benzoxazinotropones 6—10 were studied. Moreover, HMO¹⁰⁾ and MINDO/3¹¹⁾ molecular orbital theoretical calculations were performed and the theoretical results were compared with the experimental results.

a) Acylation and Methylation. No products were obtained when 6-10 were combined with acetic anhydride in pyridine at $80\,^{\circ}$ C. However, acetates 22a-d were produced in ca. 80% yields when 6-9 were mixed with acetic anhydride in acetic acid containing a trace amount of concd sulfuric acid at room temperature. Compound 22a was determined to be 6-acetoxybenzo-[b]cyclohept[e][1,4]oxazine from the high resolution mass spectral determination of molecular weight (m/z) M⁺, 253.0769) and other spectral data. The IR spec-

trum showed an O-acetyl carbonyl absorption peak near 1770 cm⁻¹. The electronic spectrum is very similar to that of the parent compound 13. The structures of 22b—d were also identified as 7-, 8-, and 9-acetoxy-benzo[b]cyclohept[e][1,4]oxazine, respectively, on the basis of their NMR and other spectral data (see the Experimental section). Similarly, 16 and 17 were reacted as above to form acetoxyl derivatives 23 and 24, respectively. Acetylated compounds 22a—d, 23, and 24 reverted to the corresponding benzoxazinotropones 6—9, 16, and 17 when treated with refluxing acetic acid for about 5 min.

Compounds 6—9 reacted slowly with diazomethane to give the corresponding methylated products 25a—d as brown needles. However, 10 did not result in acetylated and methylated compounds under similar conditions. The structures of 25a—d were assigned to 6-, 7-, 8-, and 9-methoxybenzo[b]cyclohept[e][1,4]oxazines, respectively, on the basis of their NMR and other spectral data (see the Experimental section). Methylated compounds 25a—d gradually reverted to the corresponding benzoxazinotropones 6—9 on standing in methanol at room temperature.

b) Theoretical Calculations. The resonance energies and bond currents of the keto forms 6—10 and the enol forms 6a—10a have been calculated by means of Aihara's graph theory of aromaticity. The HMO theory is assumed to be in its simplest form. Streitwieser¹⁰ evaluated the heteroatom parameters for the amine nitrogen, the

Table 1. Resonance Energies, Heats of Formation, and Circuit Resonance Energies for Keto Forms (6—10) and Enol Forms (6a—10a)

Compd	RE	$\frac{\Delta H_{\mathrm{f}}}{\mathrm{kcal\ mol^{-1}}}$	CRE		Commd	DE	$\Delta H_{ m f}$	CRE	
			r_{i}	r ₃	Compd	RE	kcal mol-1	r_1	r ₃
6	0.401	-25.1	0.152	0.216	6a	0.312	-22.2	0.126	0.201
7	0.404	-35.4	0.142	0.213	7a	0.319	-31.8	0.117	0.197
8	0.402	-29.1	0.151	0.216	8a	0.314	-28.4	0.125	0.200
9	0.403	-35.0	0.142	0.213	9a	0.318	-30.7	0.117	0.197
10	0.402	-30.0	0.152	0.216	10a	0.315	-27.2	0.130	0.201

RE: Resonance energy (in β units). CRE: Circuit resonance energy (in β units). ΔH_f : Heats of formation calculated by the MINDO/3 method.

imine nitrogen, the ether oxygen, and the ketone oxygen. We have used these values in this paper.

MINDO/3¹¹⁾ is a reasonably reliable method for the prediction of heats of formation. The molecular geometries of 6—10 and its enol forms 6a—10a were calculated by MINDO/3 optimizations. In all cases, the MINDO/3 optimized geometries were planar and exhibited appreciable bond alternation. As shown in Table 1, the heats of formation of the keto forms are more stable than those of the enol forms by 0.7—4.3 kcal mol⁻¹.

All the keto and enol compounds are predicted to be aromatic with positive resonance energies. 12a The keto form species have larger resonance energies than those of the enol forms. With a view to clarifying the origin of the aromaticity, we calculated the circuit resonance energies 12c of the keto forms by means of Aihara's graph theory of aromaticity. The π -electron ring system in the keto forms consists of six π -ring components, from r_1 to r_6 , as shown in Fig. 1. Geometrically unidentical π -electron circuits are shown and numbered in Fig. 1. The results are listed in Table 1. The circuit resonance energies of r_1 and r_3 are positive, while those of r_2 , r_4 , r_5 , and r_6 are near zero or negative. Consequently, the keto form compounds are stabilized as the 6- π -tropylium-6- π -benzenoid form.

Since 6—10 are only slightly soluble in less polar solvents, the NMR spectra were measured in DMSO- d_6 . In all cases, the 13 C NMR spectra exhibited a peak at

about δ =180 resulting from a carbonyl group. These results match the keto form but not the enol form in a polar solvent such as DMSO. In the ¹H NMR spectra, the chemical shifts of the seven-membered ring protons for 6-10 differ from those of parent compound 13. This can also be explained by assuming the compounds are in the keto form. This causes the signals of the β proton of the carbonyl groups to appear at a lower field compared with those of compounds 13, 22, and 25. The observed values of the average chemical shifts of the seven-membered ring protons and benzene ring protons of 6—10 are at δ =6.61 and 6.70, respectively. In contrast, the chemical shifts of the seven-membered ring protons of the enol forms and 13 are expected to be much higher than those of the benzene ring protons.¹³⁾ Furthermore, ring currents of the keto forms of 6-10 and their enol forms 6a-10a have been calculated by Aihara's method, 12b) and the results are shown in Fig. 1. The average ring currents of the seven-membered rings (r_1) and benzene rings (r_3) of 6—10 are 0.930 I_0 and 1.000 I_0 , respectively, whereas those of 6a-10a are $0.710 I_0$ and 1.040 I₀ respectively. We therefore conclude that benzoxazinotropones 6—10 exist mainly in the keto form on the basis of the experimental facts, NMR spectral data, and theoretical calculations.

Though compounds 6—9 were almost the same yellow-brown color and had very similar electronic absorption bands, 10 had a red color. The electronic spectra of 10, in methanol solution, showed the longest

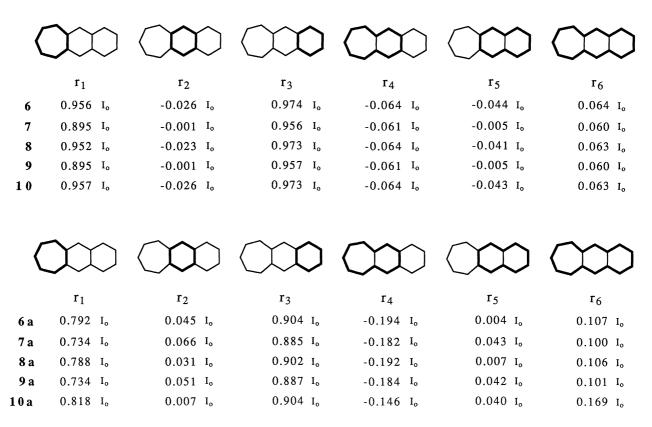


Fig. 1. Ring currents of 6—10 and 6a—10a. (I₀=Ring currents of benzene).

wavelength band centered at 480 nm. We have also calculated the electronic transitions for 6—10 by the CNDO/S-CI method. For example, the spectra of 7 showed an absorption band, deriving from the HOMO-LUMO transition, lying in the UV region at 3.13 eV (395 nm). The electronic transitions calculated by adopting the MINDO/3 geometries of 10 are predicted to have $\pi\pi^*$ $\lambda_{\rm max}$ of 2.73 eV (454 nm, f=0.000), 2.75 eV (450 nm, f=0.000), 2.98 eV (416 nm, f=0.094), and 3.05 eV (406 nm, f=0.072) in the long wavelength region. The calculated results, however, do not conform to the electronic spectra of 10.

To elucidate the molecular structure of 10, an X-ray diffraction analysis was carried out.¹⁴⁾ The molecular framework is illustrated in Fig. 2. The definitive structure determination of 10 was particularly interesting, because this compound was shown to exist in an intermolecularly hydrogen-bonded form in the solid state.

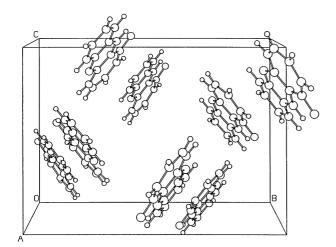


Fig. 2. A perspective view of 10.

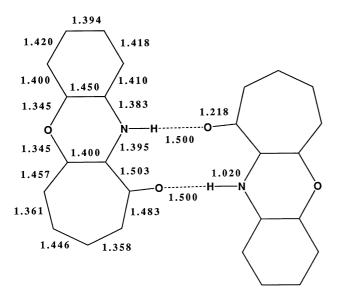


Fig. 3. Assumed structure of Model A. Bond lengths are expressed in Angstroms (Å).

The H-bond is between the ketone oxygen in one molecule and the imine hydrogen in another molecule. We therefore propose two dimeric model structures on the basis of the above X-ray experimental geometry and MINDO/3 optimized values. Model A is a planar molecule with an assumed N···O bond length of 2.718 Å (Fig. 3), whereas Model B is a nonplanar molecule. The geometries of Model B are the same as those of Model A, except that the separation of two molecular planes is assumed to be 1.9 Å and the N···O bond length is 2.2 Å. The calculated electronic transitions of Model A are predicted to have $\pi\pi^* \lambda_{\text{max}}$ of 2.57 eV (481 nm, f=0.000), 2.80 eV (442 nm, f=0.000), 2.85 eV (435 nm, f=0.166), and 2.96 eV (418 nm, f=0.238) in the long wavelength region and are very close to the calculated transitions of monomeric compound 10. The calculated electronic transitions of Model B are predicted to have $\pi\pi^* \lambda_{max}$ of 2.48 eV (500 nm, f=0.002), 2.53 eV (490 nm, f=0.215), 2.84 eV (436 nm, f=0.012), 2.92 eV (424 nm, f=0.151), 2.93 eV (423 nm, f=0.180), and 3.06 eV (405 nm, f=0.051) in the same region. In the calculated results of Model B, its next longest absorption band can be attributed to an intermolecular charge-transfer transition from one molecule to another molecule because this absorption band was assigned to a HOMO-LUMO transition, and π charges in HOMO or LUMO orbitals were distributed on each molecule of the assumed dimer. The electronic transitions calculated for Model B agree with the experimental electronic spectra of 10. agreement between calculated and experimental data is satisfactory, especially if we bear in mind that theoretical values refer to the gas phase and experimental values to the solution state.

Possible Reaction Pathways for the Formation of Benzoxazinotropones. As mentioned above, bromo compounds 11a-e gave an almost quantitative yield of benzoxazinotropones 6—10, respectively, upon heating in 97% acetic acid. However, upon heating in ethanol, 11a—d did not result in benzoxazinotropones 6—9, while 11e gave 10 almost quantitatively. Additionally, by periodically checking the reaction with reversed-phase HPLC we observed the formation of 22d when 11d was heated in anhydrous acetic acid at 110 °C. As shown in Fig. 4, a peak corresponding to acetoxyl compound 22d appears first, then the peak corresponding to 9 increases as the peaks corresponding to 22d and 11d decrease in the chromatogram. In this transformation, as shown in Scheme 4, intermediate a may be produced first by the attack of an acetoxyl anion (not by moisture in the acetic acid) at the bromine-substituted C-9 position of 11d. Compound a may then give 9 via acetoxyl derivative 22d. Similarly, 11a—c are likely to produce 6—8 via acetoxyl compounds 22a-c, respectively, and 15 is likely to produce 16 and 17 via acetoxyl compounds 23 and 24, respectively.

As we reported earlier, 7) position C-5a in the benzo [b] cyclohept [e] [1,4] oxazine nucleus is most easily

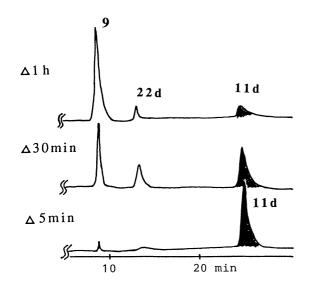


Fig. 4. Time-dependent HPLC chromatograms of the hydrolysis of **11d** in dry AcOH at 110°C.

attacked by nucleophiles, and when the C-10 position is substituted with a group such as a bromine atom, that group acts as a good leaving group. Therefore, it is quite understandable that the 10-bromo compound 11e is readily transformed to 10 even in MeOH (containing a small amount of water), and the tribromo compound 18 gives 7,9-dibromobenzoxazinotropone 19 as the main

product along with the 6,8-dibromo compound 20, as illustrated in Scheme 5.

Experimental

Melting points were determined with a Yanagimoto MP-3S melting point apparatus and were uncorrected. The IR and electronic spectra were measured by using a Shimadzu IR-450 and a Shimadzu UV-265FS spectrometer, respepctively; the UV spectra in acid and alkali solutions were taken after adding a few drops of 6 M HCl or 6 M NaOH (1 M=1 mol dm⁻³) to the sample solution. The NMR spectra were measured in CDCl₃ (unless otherwise specified) with a JEOL JNM-PS/PET (100 MHz) or a JEOL JNM-GX270 (270 MHz for 1H and 67.8 MHz for ¹³C) spectrometer using TMS as the internal standard. The assignments of all signals were made by employing a first-order analysis with the aid of a decoupling technique. The mass spectra were taken on a JEOL JMS-DX300 mass spectrometer and a Shimadzu LKB 9000 GC-mass spectrometer at 70 eV. The HPLC was carried out on Hitachi gel #3011 with MeOH-hexane (9:1) as the solvent. The centrifugal chromatography was performed with a Hitachi CLC-5 instrument in a Fuji silica-gel layer (KT-2151, 3 mm thickness) using benzene as an eluent. The TLC analyses were carried out with Merck Kieselgel 60F-254 plates.

4-Bromo-2-(2-hydroxyanilino)tropone (12c). A mixture of 5-bromotropolone p-toluenesulfonate¹⁵⁾ (14, 506 mg, 1.42 mmol) and o-aminophenol (4, 310 mg, 2.82 mmol) in pyridine (5 cm³) was heated at 60 °C for 5 h. The solvent was evaporated in vacuo and the residue was chromatographed on a column of silica gel with benzene-MeOH (50:1), giving 12c (345 mg, 84% yield):Orange needles (from MeOH); mp 170— 172 °C (decomp); UV_{max} (MeOH) 240 (log ϵ 4.20), 262 (4.31), 347 (4.04), and 407 nm (4.18), (MeOH+ 6 M NaOH) 244 (log ε 4.29), 272 (4.17), 346 (3.89), and 413 nm (4.11); IR (KBr) 3430 (OH) and 3250 cm⁻¹ (NH); ¹H NMR (270 MHz, CD₃OD) δ =6.96 (1H, td, J=8.0 and 2.0 Hz, H-5'), 7.00 (1H, dd, J=8.0 and 2.0 Hz, H-3'), 7.05—7.10 (4H, m, H-3,5,6,7), 7.20 (1H, td, J=8.0 and 2.0 Hz, H-4'), and 7.28 (1H, dd, J=8.0 and 2.0 Hz, H-6'). Found: C, 53.26; H, 3.57; N, 4.60%; M+, 292. Calcd for C₁₃H₁₀NO₂Br: C, 53.45; H, 3.45; N, 4.79%; M, 292.

9-Bromobenzo[b]cyclohept[e][1,4]oxazine (11d). A solution of 12c (210 mg) in acetic acid (5 cm³) containing a trace amount of concd sulfuric acid was heated at 120 °C for 2 h under nitrogen. The solvent was then evaporated in vacuo. The residue was neutralized with aq NaHCO₃ and extracted twice with benzene. The extracts were combined, dried (Na₂SO₄) and evaporated in vacuo. The residue was chromatographed centrifugally on a layer of silica gel with benzene as the eluent, giving 11d (161 mg, 82% yield): Brown needles; mp 145—147 °C (from benzene); UV $_{max}$ (MeOH) 263 (log ϵ 4.47), 270 (4.47), 305 (4.03), and 414 nm (4.10), (MeOH +6M HCl) 267 (log ϵ 4.37), 287 (4.27), 320 (4.15), and 449 nm (3.99); ${}^{1}H$ -NMR (270 MHz) δ =5.29 (1H, d, J=9.8 Hz, H-6), 5.70 (1H, dd, J=11.5 and 9.8 Hz, H-7), 5.90 (1H, dd, J=11.5 and 2.0 Hz, H-8), 6.40 (1H, m, H-4), 6.54 (1H, d, J=2.0 Hz, H-10), and 6.78 (3H, m, H-1,2,3). Found: C, 57.00; H, 3.08; N, 4.99%; M+, 274. Calcd for C₁₃H₈NOBr: C, 56.96; H, 2.94; N, 5.11%; M, 274.

7-Bromobenzo[b]cyclohept[e][1,4]oxazine (11b). A mixture of 11d (30 mg, 0.11 mmol) and 4 (24 mg, 0.22 mmol) in methanol (5 cm³) was allowed to stand for 1 h at room

temperature. The reaction contents were checked by means of time-dependent HPLC. The solvent was evaporated in vacuo. The residue was chromatographed on silica-gel thin-layer plates with benzene as the eluent. The product was recrystallized from benzene, giving 11b (13 mg, 45% yield)(along with 50% recovered 11d): Brown needles; mp 130—132 °C; UV_{max} (MeOH) 222 (log ε 4.31), 262 (4.39), 272 (4.36), 309 (4.15 sh), 322 (3.76 sh), and 415 nm (4.05), (MeOH+6 M HCl) 230 (log ε 4.35), 267 (4.30), 285 (4.17 sh), 317 (4.01), 439 (3.98), and 427 nm (3.87 sh); 1 H NMR (270 MHz) δ =5.69 (1H, d, J=1.5 Hz, H-6), 5.82 (1H, dd, J=12.4 and 8.8 Hz, H-9), 6.00 (1H, d, J=12.4 Hz, H-10), 6.12 (1H, dd, J=8.8 and 1.5 Hz, H-8), 6.43 (1H, m, H-4), and 6.73—6.85 (3H, m, H-1,2,3). Found: C, 56.74; H, 3.05; N, 4.89%; M+, 274. Calcd for C₁₃H₈NOBr: C, 56.96; H, 2.94; N, 5.11%; M, 274.

Similarly, when a methanolic solution of 11b and 4 was allowed to stand at room temperature, an additional peak due to compound 11d began to appear in the HPLC chromatogram and an equilibrium was reached within a few hours. The isomerized 11d was isolated in pure crystalline form.

Hydrolysis of 11a—e. A solution of 11c (55 mg) in acetic acid (10 cm³) was heated at 110 °C for 1 h, followed by the same procedures as those described above for 12c. The residual solid was recrystallized from MeOH to give 8 (40 mg, 95% yield). Similarly, 6⁴ (95% yield), 7 (95% yield), 9 (90% yield), and 10⁴ (95% yield) were obtained from 11a, 11b, 11d, and 11e, respectively.

Benzo[*b*]cyclohept[*e*][1,4]oxazin-6(11*H*)-one (6): Mp 270—271 °C (lit, ⁴) mp 270 °C); UV_{max} (DMSO) 264, 305, 315, 330 (sh), and 393 nm, (CHCl₃) 259, 300, 310, 326 (sh), and 388 nm; ¹H NMR (270 MHz, DMSO- d_6) δ=6.42 (1H, dd, J=8.0 and 1.8 Hz, H-4), 6.49 (1H, dd, J=12.2 and 2.6 Hz, H-7), 6.56 (1H, dd, J=8.0 and 1.8 Hz, H-1), 6.64 (1H, td, J=8.0 and 1.8 Hz, H-2), 6.72 (1H, td, J=8.0 and 1.8 Hz, H-3), 6.81 (3H, m, H-8,9,10), and 8.57 (1H, br, NH); ¹³C NMR (67.8 MHz, DMSO- d_6) δ=113.4 (d), 114.9 (d), 123.1 (d), 124.6 (d), 126.9 (d), 130.9 (s), 131.0 (d), 131.6 (d), 139.9 (s), 140.9 (d), 143.2 (s), 145.2 (s), and 173.3 (s).

Benzo[*b*]cyclohept[*e*][1,4]oxazin-7(11*H*)-one (7): Orange needles; mp 294—296 °C (from MeOH); UV_{max} (MeOH) 221 (log ε 4.27), 237 (4.14), 295 (4.29), 325 (3.70 sh), and 427 nm (4.02), (MeOH+6 M HCl) 225 (log ε 4.30), 256 (4.14), 287 (4.20), 309 (3.99 sh), and 452 nm (3.91), (MeOH+6 M NaOH) 258 (log ε 4.28), 300 (4.25), 465 (4.17), and 497 nm (4.08), (DMSO) 265, 289, 396 (sh), and 423 nm, (CHCl₃) 242, 288, 296, 390 (sh), and 413 nm; IR (KBr) 3250 and 1640 cm⁻¹; ¹H NMR (270 MHz, DMSO- d_6) δ=5.99 (1H, d, J=9.7 Hz, H-10), 6.13 (1H, dd, J=11.8 and 2.7 Hz, H-8), 6.33 (1H, d, J=2.7 Hz, H-6), 6.70 (1H, dd, J=8.0 and 1.8 Hz, H-4), 6.75 (1H, td, J=8.0 and 1.8 Hz, H-2), 6.81 (1H, dd, J=11.8 and 9.7 Hz, H-9), 6.84 (1H, dd, J=8.0 and 1.8 Hz, H-1), 6.89 (1H, td, J=8.0 and 1.8 Hz, H-3), and 9.93 (1H, br, NH). Found: m/z 211.0656. Calcd for C₁₃H₉NO₂: M, 211.0633.

Benzo[*b*]cyclohept[*e*][1,4]oxazin-8(11*H*)-one (8): Brown needles; mp>300 °C (from MeOH); UV_{max} (MeOH) 222 (log ε 4.12), 266 (4.34), 275 (4.32), 318 (3.39 sh), and 420 nm (4.02), (MeOH+6 M HCl) 227 (log ε 4.25), 266 (4.29), 275 (4.31), 330 (3.60), 443 (3.81), and 466 nm (3.79 sh), (DMSO) 268, 277, 315, and 408 nm, (CHCl₃) 264, 274, 320 (sh), and 402 nm; IR (KBr) 3250—2800 and 1644 cm⁻¹; ¹H NMR (270 MHz, DMSO- d_6) δ=6.36 (1H, dd, J=12.7 and 2.0 Hz, H-7), 6.43 (1H, dd, J=8.0 and 2.0 Hz, H-4), 6.52 (1H, dd, J=8.0 and 2.0 Hz, H-1), 6.64

(1H, td, J=8.0 and 2.0 Hz, H-3), 6.69 (2H, m, H-9,10), 6.73 (1H, td, J=8.0 and 2.0 Hz, H-2), 6.75 (1H, d, J=12.7 Hz, H-6), and 8.70 (1H, br, NH); ¹H NMR (270 MHz, CD₃OD) δ =6.57 (1H, dd, J=13.1 and 2.5 Hz, H-7), 6.77 (1H, d, J=13.1 Hz, H-10), 6.84 (1H, dd, J=13.1 and 2.5 Hz, H-9), and 6.89 (1H, d, J=13.1 Hz, H-6); ¹³C NMR (67.8 MHz, DMSO- d_6) δ =114.3 (d), 114.9 (d), 123.7 (d), 124.9 (d), 129.5 (d), 129.6 (d), 130.0 (s), 131.1 (d), 138.8 (d), 139.7 (s), 143.2 (s), 143.4 (s), and 179.5 (s). Found: C, 73.76; H, 3.99; N, 6.28%; M+, 211. Calcd for C₁₃H₉NO₂: C, 73.92; H, 4.29; N, 6.63%; M, 211.

Benzo[b]cyclohept[e][1,4]oxazin-9(11H)-one (9): Yellow needles; mp 243-245 °C (from MeOH); UV_{max} (MeOH) 239 $(\log \varepsilon 4.19)$, 294 (4.48 sh), 307 (4.55), 338 (4.00 sh), 373 (3.77), and 394 nm (3.77), (MeOH+6 M HCl) 226 (log ε 4.14), 251 (4.22), 257 (4.24), 289 (4.36), 308 (4.54), and 426 nm (3.94), (MeOH+6 M NaOH) 240 (log ε 3.99), 305 (4.54), 324 (4.27 sh), 340 (4.07 sh), 423 (4.09), and 453 nm (3.91 sh), (DMSO) 298, 308, 340 (sh), and 380 nm (sh), (CHCl₃) 296, 307, 340 (sh), and 394 nm (sh); IR (KBr) $3250 \text{ and } 1614 \text{ cm}^{-1}$; ^{1}H NMR (270 MHz, DMSO- d_6) δ =6.05 (1H, d, J=2.5 Hz, H-10), 6.32 (1H, dd, J=12.4 and 2.5 Hz, H-8), 6.39 (1H, d, J=10.3 Hz, H-6), 6.70 (1H, dd, J=12.4 and 10.3 Hz, H-7), 6.71 (1H, dd, J=8.0 and 1.5 Hz, H-4), 6.78 (1H, td, J=7.5 and 1.5 Hz, H-2), 6.83 (1H, dd, J=8.0 and 1.5 Hz, H-1), 6.88 (1H, td, J=7.5 and 1.5 Hz, H-3), and 9.80 (1H, br, NH); ¹³C NMR (67.8 MHz, DMSO-d₆) δ =114.0 (d), 114.2 (d), 114.8 (d), 115.2 (d), 121.8 (d), 124.2 (d), 127.1 (s), 132.2 (d), 136.3 (d), 140.8 (s), 141.5 (s), 151.0 (s), and 183.1 (s). Found: C, 73.61; H, 4.38; N, 6.30%; M+, 211. Calcd for $C_{13}H_9NO_2$: C, 73.92; H, 4.29; N, 6.63%; M, 211.

Benzo[b]cyclohept[e][1,4]oxazin-10(11H)-one (10): Mp 171-172 °C (lit, 4) mp 170 °C); UV_{max} (DMSO) 265, 273, 312, 330, 406, and 487 nm, (CHCl₃) 261, 271, 305, 324, 404, and 483 nm; ¹H NMR (270 MHz, DMSO- d_6) δ =6.57 (1H, dd, J=8.0 and 2.0 Hz, H-4), 6.68 (2H, m, H-3,7), 6.73 (1H, td, J=8.0 and 2.0 Hz, H-2), 6.78 (1H, dd, J=11.1 and 1.2 Hz, H-6), 6.93 (1H, dd, J=8.0 and 2.0 Hz, H-1), 6.94 (1H, dd, J=11.4 and 1.5 Hz, H-9), 7.03 (1H, ddd, J=11.4, 8.3, and 1.5 Hz, H-8), and 8.85 (1H, br, NH); ¹³C NMR (67.8 MHz, DMSO- d_6) δ =114.4 (d), 115.5 (d), 123.8 (d), 124.6 (d), 125.2 (d), 128.2 (d), 129.3 (s), 133.9 (d), 134.2 (d), 141.4 (s), 143.7 (s), 146.5 (s), and 171.9 (s).

Acetylation of 6—10. A mixture of 9 (10 mg, 0.05 mmol) and acetic anhydride (0.6 cm³) in acetic acid (1 cm³) containing a trace amount of concd sulfuric acid was stirred for 2 h at room temperature, followed by the same procedures as those described above for 11b. The residual solid was recrystallized from MeOH to give 22d (10 mg, 83% yield). Similarly, 22a (80% yield), 22b (80% yield), and 22c (80% yield) were obtained from 6, 7, and 8, respectively. Compound 10 was recovered unchanged under similar conditions.

6-Acetoxybenzo[*b*]**cyclohept**[*e*][1,4]**oxazine** (22a): Brown needles; mp 120—122 °C (from MeOH); UV_{max} (MeOH) 259 (log ε 4.33), 267 (4.30), 300 (3.80), 316 (3.70 sh), 330 (3.54 sh), and 405 nm (3.99), (MeOH+6 M HCl) 225 (log ε 4.35), 263 (4.29), 271 (4.33), 319 (3.84), 431 (3.91), and 453 nm (3.88 sh); IR (KBr) 1770 cm⁻¹ (C=O); ¹H NMR (270 MHz, CD₃CN) δ=2.19 (3H, s, Me), 5.89 (1H, ddd, J=12.4, 6.4, and 2.5 Hz, H-8), 5.98 (1H, dd, J=12.4 and 2.0 Hz, H-7), 6.19 (1H, dd, J=12.5 and 2.5 Hz, H-10), 6.23 (1H, ddd, J=12.5, 6.4, and 2.0 Hz, H-9), 6.50 (1H, m, H-4), and 6.82 (3H, m, H-1,2,3). Found: C, 70.92; H, 4.65; N, 5.34%; M⁺, 253. Calcd for C₁₅H₁₁NO₃: C, 71.14; H, 4.38; N, 5.53%; M, 253.

7-Acetoxybenzo[b]cyclohept[e][1,4]oxazine (22b); Brown

needles; mp 95—97 °C (from MeOH); UV_{max} (MeOH) 259 (log ϵ 4.17), 268 (4.12), 294 (3.95), and 410 nm (3.94), (MeOH+6 M HCl) 225 (log ϵ 4.27), 261 (4.17), 272 (4.12), 311 (3.82), 429 (3.77), and 457 nm (3.75 sh); IR (KBr) 1755 cm⁻¹ (C=O); ¹H NMR (270 MHz, CD₃CN) δ =2.14 (3H, s, Me), 5.60 (1H, d, J=2.0 Hz, H-6), 5.84 (1H, dd, J=10.0 and 2.0 Hz, H-8), 6.37 (1H, dd, J=12.0 and 10.0 Hz, H-9), 6.47 (2H, m, H-4,10), and 6.80 (3H, m, H-1,2,3). Found: m/z 253.0737. Calcd for C₁₅H₁₁NO₃: M, 253.0737.

8-Acetoxybenzo[b]cyclohept[e][1,4]oxazine (22c): Brown needles; mp 139—141 °C (from MeOH); UV_{max} (MeOH) 260 (log ϵ 4.36), 269 (4.32), 300 (3.82), 320 (3.70 sh), 334 (3.66 sh), and 415 nm (4.05), (MeOH+6 M HCl) 225 (log ϵ 4.35), 263 (4.38), 272 (4.41), 323 (3.87), 441 (3.97), and 467 nm (3.93 sh); IR (KBr) 1750 cm⁻¹ (C=O); ¹H NMR (270 MHz, CD₃CN) δ =2.10 (3H, s, Me), 5.40 (1H, d, J=10.1 Hz, H-6), 5.77 (1H, dd, J=10.1 and 2.7 Hz, H-7), 6.00 (1H, dd, J=12.7 and 2.7 Hz, H-9), 6.10 (1H, d, J=12.7 Hz, H-10), 6.42 (1H, m, J=8.0 and 2.0 Hz, H-4), and 6.77 (3H, m, H-1,2,3). Found: m/z 253.0721. Calcd for C₁₅H₁₁NO₃: M, 253.0737.

9-Acetoxybenzo[*b*]**cyclohept**[*e*][1,4]**oxazine** (22d): Brown needles; mp 149—151 °C (from MeOH); UV_{max} (MeOH) 260 (log ε 4.49), 268 (4.39), 295 (3.94), and 410 nm (4.07), (MeOH+6 M HCl) 226 (log ε 4.29), 262 (4.32), 272 (4.37), 315 (4.00), 432 (3.95), and 457 nm (3.91 sh); IR (KBr) 1743 cm⁻¹ (C=O); ¹H NMR (270 MHz, CD₃CN) δ=2.12 (3H, s, Me), 5.40 (1H, d, J=9.6 Hz, H-6), 5.57 (1H, dd, J=11.6 and 2.6 Hz, H-8), 5.79 (1H, d, J=2.6 Hz, H-10), 6.02 (1H, dd, J=11.6 and 9.6 Hz, H-7), 6.44 (1H, dd, J=8.0 and 2.0 Hz, H-4), and 6.66—6.88 (3H, m, H-1,2,3). Found: C, 70.85; H, 4.55; N, 5.29%; M⁺, 253. Calcd for C₁₅H₁₁NO₃: C, 71.14; H, 4.38; N, 5.53; M, 253.

Methylation of 6—10. Ethereal diazomethane (0.5 mmol) was added to a solution of 9 (10 mg, 0.05 mmol) in ether (2 cm³) and the mixture was stirred for 24 h at room temperature. After removal of the ether, the residue was recrystallized from benzene to give 25d (10 mg, 80% yield). Similarly, 25a (70% yield), 25b (70% yield), and 25c (80% yield) were obtained from 6, 7, and 8, respectively. Compound 10 was recovered unchanged under similar conditions.

6-Methoxybenzo[b]cyclohept[e][1,4]oxazine (25a): Brown needles; mp 94—96 °C (from benzene); UV_{max} (MeOH) 263, 269, 312, and 415 nm, (MeOH+6 M HCl) 230, 275, 310, and 436 nm; ¹H NMR (270 MHz, CD₃CN) δ =3.68 (3H, s, OMe), 6.08 (1H, ddd, J=12.0, 6.5, and 3.0 Hz, H-8), 6.34 (2H, m, H-7,9), 6.39 (1H, d, J=12.0 Hz, H-10), 6.49 (1H, m, H-4), and 6.74—6.85 (3H, m, H-1,2,3). Found: m/z 225.0771. Calcd for C₁₄H₁₁NO₂: M, 225.0790.

7-Methoxybenzo[*b*]**cyclohept**[*e*][**1,4**]**oxazine** (**25b**): Brown needles; mp 87—89 °C (from benzene); UV_{max} (MeOH) 222, 260, 268, 325, and 414 nm, (MeOH+6 M HCl) 226, 261, 271, 310, 430, and 467 nm (sh); ¹H NMR (270 MHz) δ =3.66 (3H, s, OMe), 5.65 (1H, d, J=1.5 Hz, H-6), 5.92 (1H, dd, J=12.5 and 9.7 Hz, H-9), 6.10 (1H, d, J=12.5 Hz, H-10), 6.25 (1H, dd, J=9.7 and 1.5 Hz, H-8), 6.49 (1H, m, H-4), and 6.74—6.86 (3H, m, H-1,2,3); MS (70 °C) m/z 225 (M⁺, 64%), 210 (M⁺—Me, 24%), and 182 (M⁺—Me—CO, 100%). Found: m/z 225.0763. Calcd for C₁₄H₁₁NO₂: M, 225.0790.

8-Methoxybenzo[*b***]cyclohept[***e***][1,4]oxazine (25c):** Brown needles; mp 105—107 °C (from benzene); UV_{max} (MeOH) 220, 263, 272, 320, and 427 nm, (MeOH+6 M HCl) 228, 265, 271, 300, 325, 440, and 510 nm (sh); 1 H NMR (270 MHz) δ =3.58 (3H, s, OMe), 5.35 (1H, dd, J=10.3 and 2.0 Hz, H-7), 5.51 (1H,

d, J=10.3 Hz, H-6), 6.12 (1H, dd, J=13.0 and 2.0 Hz, H-9), 6.28 (1H, d, J=13.0 Hz, H-10), 6.34 (1H, dd, J=7.5 and 1.5 Hz, H-4), and 6.64—6.81 (3H, m, H-1,2,3). Found: m/z 225.0793. Calcd for $C_{14}H_{11}NO_2$: M, 225.0790.

9-Methoxybenzo[*b*]cyclohept[*e*][1,4]oxazine (25d): Brown needles; mp 89—91 °C (from benzene); UV_{max} (MeOH) 224 (log ε 4.09), 258 (4.39), 267 (4.44), 350 (3.99 sh), 412 (4.08), and 448 nm (3.72 sh), (MeOH+6 M HCl) 225 (log ε 4.13), 252 (4.25), 259 (4.30), 287 (4.32), 305 (4.21), and 432 nm (3.94); ¹H NMR (270 MHz) δ=3.64 (3H, s, OMe), 5.44 (1H, d, J=9.7 Hz, H-6), 5.69 (2H, m, H-8,10), 5.99 (1H, dd, J=12.3 and 9.7 Hz, H-7), 6.47 (1H, m, H-4), and 6.72—6.86 (3H, m, H-1,2,3). Found: C, 74.35; H, 5.06; N, 5.93%; M⁺, 225. Calcd for C₁₄H₁₁NO₂: C, 74.65; H, 4.92; N, 6.22%; M, 225

Hydrolysis of 22a-d. Similar procedures as those described above for compounds **11a-e** were followed. Thus, the treatment of **22a-d** (ca. 5 mg) with AcOH (2 cm³) at 100 °C for 5—15 min gave **6**—9, respectively, in an almost quantitative yield.

Hydrolysis of 25a—d. Solutions of 25a—d (ca. 2 mg) in methanol (5 cm³) were allowed to stand for 48 h at room temperature. After removal of the solvent in vacuo, the residue was recrystallized from methanol, giving 6—9 in an almost quantitative yield.

Hydrolysis of 15. A solution of 15% (50 mg) in acetic acid (10 cm³) was heated at 110 °C for 10 h. The reaction mixture was chromatographed over thin-layer plates with benzene–MeOH (10:1) as the eluent, giving 16 (2 mg, 5% yield), 17 (34 mg, 80% yield), and 12d% (trace).

8-Bromobenzo[*b*]**cyclohept**[*e*][1,4]**oxazin-6(11***H*)**-one** (16): Brown solid; UV_{max} (MeOH) 230, 260, 300, 320 (sh), and 400 nm; MS (70 °C): m/z 291 (M⁺, 15%), 289 (M⁺, 17%), 263 (M⁺—CO, 71%), 261 (M⁺—CO, 65%), 182 (67%), and 154 (100%). Found: m/z 290.9741 and 288.9739 (1:1). Calcd for C₁₃H₈NO₂Br: M, 290.9718 and 288.9738.

6-Bromobenzo[b]cyclohept[e][1,4]oxazin-8(11H)-one (17): Brown solid; UV_{max} (MeOH) 250, 275, 315, and 410 nm, (MeOH+6 M HCl) 236, 264, 270, 320, and 435 nm, (MeOH+6 M NaOH) 238, 265, 275, 320, 463, and 495 nm (sh). Found: m/z 290.9759 and 288.9741 (1:1). Calcd for $C_{13}H_8NO_2Br$: M, 290.9718 and 288.9738.

Acetylation of 16 and 17. Acetylation of 16 and 17 was conducted in a manner similar to that described above for 6—10, and gave 23 and 24, respectively, in an almost quantitative yield.

6-Acetyloxy-8-bromobenzo[*b*]**cyclohept**[*e*][**1,4**]**oxazine (23):** Brown needles; mp 139—140 °C (from benzene); UV_{max} (MeOH) 220 (log ε 4.33), 258 (4.40), 269 (4.32), 294 (3.94), 305 (3.94), 319 (3.89 sh), and 409 nm (4.08), (MeOH+6 M HCl) 232 (log ε 4.29), 263 (4.40), 270 (4.42), 325 (3.95), 445 (4.03), and 472 nm (3.98 sh); IR (KBr) 1770 cm⁻¹ (C=O); ¹H NMR (270 MHz) δ=2.22 (3H, s, Me), 6.05 (1H, d, J=13.0 Hz, H-10), 6.27 (1H, d, J=2.0 Hz, H-7), 6.28 (1H, dd, J=13.0 and 2.0 Hz, H-9), 6.41 (1H, m, H-4), and 6.76—6.90 (3H, m, H-1,2,3). Found: m/z 330.9792 and 332.9832 (1:1). Calcd for C₁₅H₁₀NO₃Br: M, 330.9844 and 332.9824.

8-Acetyloxy-6-bromobenzo[*b*]**cyclohept**[*e*][1,4]**oxazine**(24): Brown needles; mp 110—111 °C (from benzene); UV_{max} (MeOH) 259 (log ε 4.25), 268 (4.23), 307 (3.76), 320 (3.71 sh), and 410 nm (3.90), (MeOH+6 M HCl) 227 (log ε 4.27), 263 (4.42), 270 (4.24), 321 (3.81), 441 (3.82), and 471 nm (3.77 sh); IR (KBr) 1750 cm⁻¹ (C=O); ¹H NMR (270 MHz) δ =2.17 (3H, s,

Me), 5.89 (1H, dd, J=13.0 and 2.2 Hz, H-9), 6.18 (1H, d, J=13.0 Hz, H-10), 6.22 (1H, d, J=2.2 Hz, H-7), 6.60 (1H, dd, J=8.0 and 2.0 Hz, H-4), 6.78—6.94 (3H, m, H-1,2,3). Found: C, 54.50; H, 3.27; N, 3.95%; M⁺, 332. Calcd for C₁₅H₁₀NO₃Br: C, 54.24; H, 3.03; N, 4.22%; M, 332.

Hydrolysis of 18. A solution of 18% (25 mg) in acetic acid (5 cm³) was heated at 110 °C for 5 h, followed by the same procedures as those described above for 15, and produced 19 (13 mg, 60% yield) and 20 (4 mg, 20% yield).

7,9-Dibromobenzo[*b*]**cyclohept**[*e*][**1,4**]**oxazin-10(11H)-one** (**19)**: Reddish-violet needles; mp 256—257 °C (from benzene); UV_{max} (MeOH) 256 (log ε 4.42), 272 (4.26 sh), 302 (4.20), 339 (3.79 sh), 418 (3.51 sh), and 520 nm (4.08), (MeOH+6 M NaOH) 260 (log ε 4.31), 274 (4.25), 304 (4.16), and 503 nm (4.16); IR (KBr) 3270 and 1557 cm⁻¹; ¹H NMR (100 MHz) δ =6.43—6.82 (4H, m, H-1,2,3,4), 7.62 (1H, d, J=2.0 Hz, H-6), and 8.08 (1H, d, J=2.0 Hz, H-8); MS (140 °C): m/z 370 (M+, 50%), 368 (M+, 100%), and 366 (M+, 51%). Found: C, 42.41; H, 2.12; N, 3.59%. Calcd for C₁₃H₇NO₂Br₂: C, 42.31; H, 1.91; N, 3.80%. Compound **19** was also obtained (90 mg, 90% yield) by heating **21**¹⁶) (100 mg, 0.27 mmol) with **4** (50 mg, 0.46 mmol) in ethanol (3 cm³) at 60 °C for 2 h, followed by the same procedures as those described above for **12c**.

6,8-Dibromobenzo[*b*]**cyclohept**[*e*][**1,4**]**oxazin-10(11***H***)-one** (**20)**: Reddish-violet solid; UV_{max} (MeOH) 225, 260, 270 (sh), 320, 420, and 510 nm; 1 H NMR (270 MHz) δ =6.54 (2H, m, H-2,4), 6.77 (2H, m, H-1,3), 7.12 (1H, d, J=1.8 Hz, H-9), 7.84 (1H, br, NH), and 8.08 (1H, d, J=1.8 Hz, H-7). Found: m/z 370.8789, 368.8847, and 366.8838 (1:2:1). Calcd for C₁₃H₇NO₂Br₂: M, 370.8804, 368.8824, and 366.8844.

Reduction of 19 and 20 Zinc dust (20 mg) was added to a solution of 19 (30 mg, 0.08 mmol) in acetic acid (5 cm³), and the mixture was stirred at room temperature for 2 h. The reaction mixture was filtered and evaporated to dryness in vacuo. The residue was then chromatographed on silica-gel thin layer plates with benzene–MeOH (50:1) as the eluent, giving compound 10⁴) (40% yield). Compound 10 was also obtained from 20 by following an identical procedure.

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References

- 1) Part III: T. Nozoe, H. Okai, H. Wakabayashi, and S. Ishikawa, *Bull. Chem. Soc. Jpn.*, **62**, 2307 (1989).
- 2) Part of these results have been preliminarily reported: H. Wakabayashi, S. Ishikawa, H. Okai, and T. Nozoe, The 50th National Meeting of the Chemical Society of Japan, Tokyo, April 1985, Abstr., 1T16 and 1T17.
- 3) T. Nozoe and T. Someya, *Bull. Chem. Soc. Jpn.*, **51**, 3316 (1978).
- 4) T. Someya, H. Okai, H. Wakabayashi, and T. Nozoe, Bull. Chem. Soc. Jpn., 56, 2756 (1983).
- 5) T. Nozoe, T. Someya, and H. Okai, *Bull. Chem. Soc. Jpn.*, **52**, 1156 (1979).
- 6) H. Wakabayashi, S. Ishikawa, H. Okai, and T. Nozoe, Bull. Chem. Soc. Jpn., 58, 2840 (1985); T. Nozoe, H. Wakabayashi, and S. Ishikawa, Heterocycles, 29, 1005 (1989).
- 7) T. Nozoe, H. Okai, H. Wakabayashi, and S. Ishikawa, Chem. Lett., 1988, 1589.
- 8) T. Nozoe, K. Takase, H. Matsumura, T. Asao, K. Kikuchi, and S. Itô, "Dai Yuki Kagaku," ed by M. Kotake, Asakura Shoten, Tokyo (1960), Vol. XIII.
- 9) T. Nozoe, K. Shindo, H. Wakabayashi, and S. Ishikawa, Heterocycles, 29, 1459 (1989).
- 10) A. Streitwieser, Jr., "Molecular Orbital Theory for Organic Chemists," Wiley, New York (1961).
- 11) J. J. P. Stewart and F. J. Seiler, MOPAC QCPE #549.
- 12) a) J. Aihara, J. Am. Chem. Soc., 98, 2750 (1976); b) ibid., 107, 298 (1985); c) ibid., 99, 2048 (1977).
- 13) T. Nozoe, H. Okai, and T. Someya, *Bull. Chem. Soc. Jpn.*, **51**, 2185 (1978).
- 14) The crystal data of **10** were as follows: $C_{11}H_9NO_2$, monoclinic with the space group $P2_1/c$, a=10.320(1), b=16.333(5), c=11.937(4) Å, $\beta=97.71(3)^\circ$, V=1994(1) ų, Z=8, Dc=1.411 g cm⁻³. 3334 independent reflections $(2\theta<126^\circ$; $|F_o|>3\sigma|F_o|)$ were collected with a Rigaku AFC-5 automatic diffractometer, using graphite-monochromated Cu K_α radiation. The final R value was 0.0620. The molecular framework was illustrated; C. Kabuto, J. Okada, A. Kawamata, H. Wakabayashi, S. Ishikawa, and T. Nozoe, to be published elsewhere.
- 15) W. von E. Doering and C. F. Hiskey, *J. Am. Chem. Soc.*, **74**, 5688 (1952).
- 16) K. Kikuchi, Bull. Chem. Soc. Jpn., 33, 628 (1960).